

PREVALENCE OF ENDOPARASITES  
IN PET FERRETS AND SMALL RODENTS



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**Introduction**

There are limited studies on the prevalence of endoparasites in pet rodents and ferrets. Most are older<sup>1,2,3,5,6</sup>, evaluate only low case numbers and relate only to certain parasite species<sup>2</sup> or animal species<sup>1,6,7</sup>. Therefore, endoparasite results from faecal samples of 1408 ferrets and small rodents examined in a diagnostic laboratory (2019-2022) were evaluated.

**Patients and Methods**

- Faecal samples from 416 ferrets, 305 rats, 180 gerbils, 173 mice, 117 hamsters, 114 chinchillas and 103 degus were examined for endoparasites.
- 1280 of the samples originated from German-speaking countries (Germany n=1157, Austria n=98, Switzerland n=25) and 128 samples from other European countries (Spain, Romania, Sweden, Denmark, Finland, Norway, France, Italy, Slovakia, Luxembourg, Netherlands, Bulgaria, Czech Republic, Belgium, Slovenia).
- The faecal samples were examined microscopically after enrichment using the flotation method (salt-glucose solution, specific gravity 1.3) and the sodium acetate-acetic acid-formalin concentration (SAFC).

**Results**

- The detection rate of parasites in ferrets was 3.8% and in small rodents between 14.6% and 27.8% (Fig. 1).
- In 15.2% of the positive samples (n=237), several parasite species were present per sample.
- Ferrets
  - Coccidian oocysts of the Eimeriidae family were most frequently detected in ferrets, followed by hookworm eggs and *Crenosoma* larvae (Fig. 2).
- Rodents
  - In all 6 small rodent species *Giardia* spp. cysts were microscopically detectable.
  - The highest number of parasite species was found in rats (n=10), followed by gerbils (n=8). In chinchillas only one parasite species was found (Tab. 1).
  - Eight of eleven cestode eggs were further differentiated as *Hymenolepis nana*.

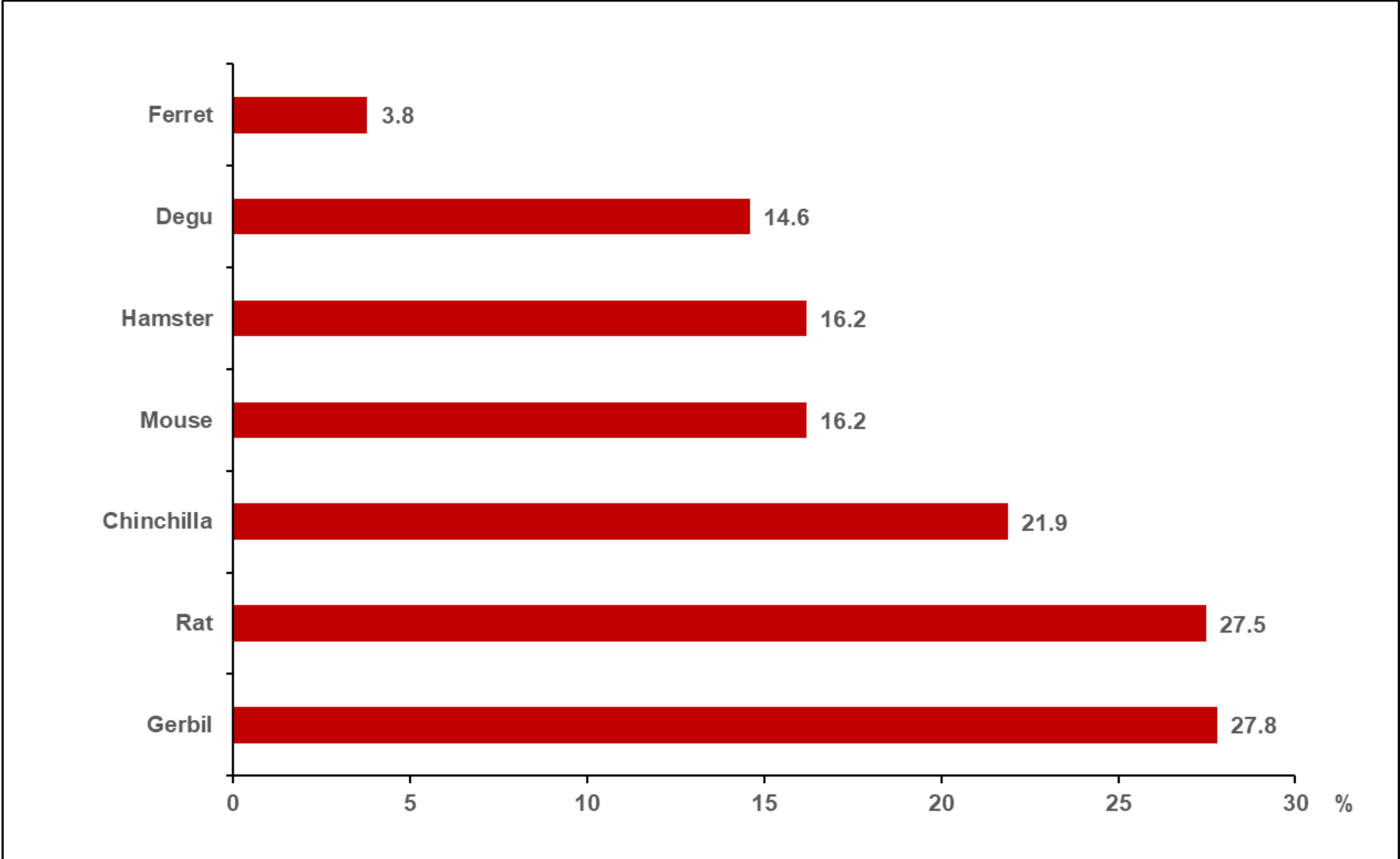


Fig. 1. Frequency of positive faecal samples for parasites in ferrets and small rodents in the years 2019-2022 (%)

Tab. 1. Detection rate of parasite stages in faecal samples of small rodents in the years 2019-2022 (%)

	Rat	Mouse	Gerbil	Hamster	Degu	Chinchilla
<i>Giardia</i> spp.	13.1	2.9	3.3	6.8	10.7	21.9
<i>Eimeria</i> spp.	3.0	1.2	3.9		1.9	
Oxyurids	10.5	13.9	21.1	9.4	2.9	
Cestodes	2.0	1.2	0.6	1.7		
Strongylids	1.3		1.1			
<i>Capillaria</i> -like eggs	0.7		0.6	4.3		
<i>Heterakis</i> spp.	0.7		1.1			
<i>Trichuris muris</i>	0.3	0.6				
<i>Strongyloides ratti</i>	0.3					
<i>Entamoeba muris</i>	0.3		1.1			

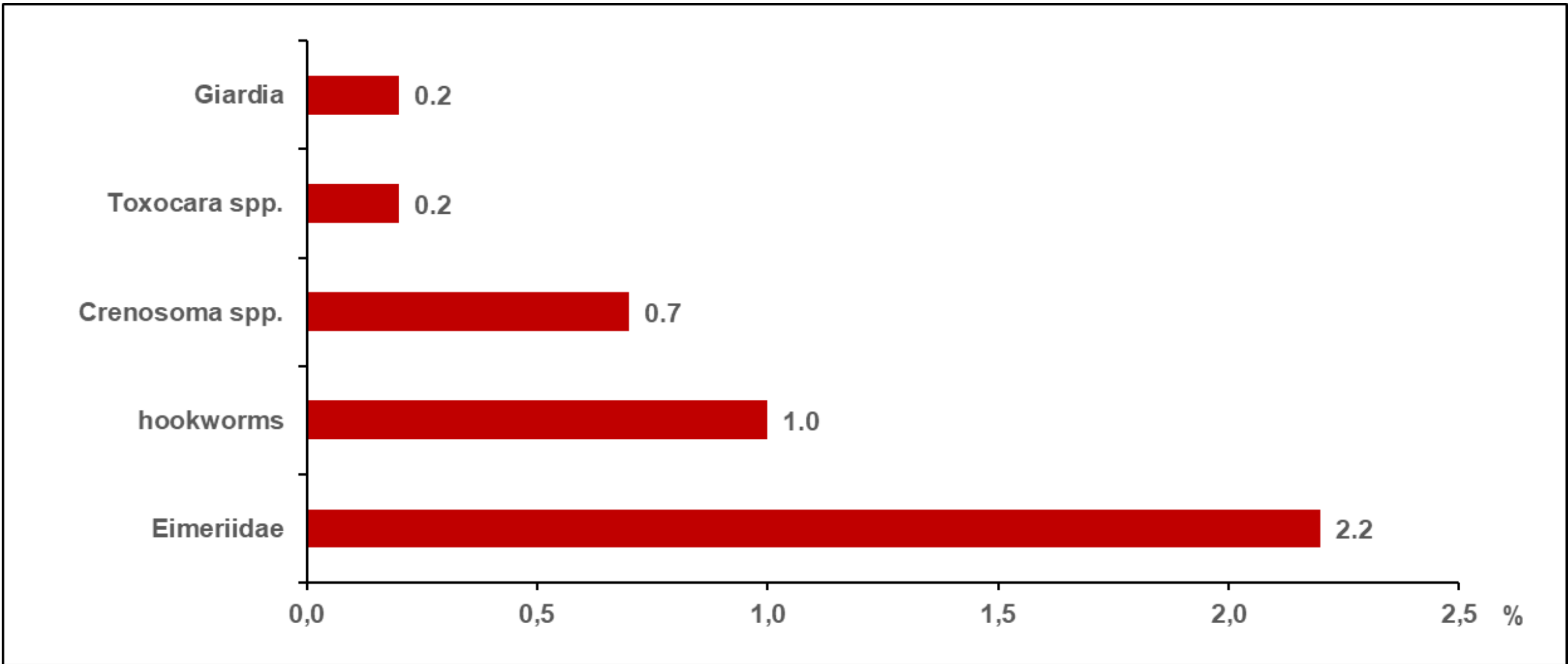


Fig. 2. Detection rate of parasite stages in faecal samples of ferrets in the years 2019-2022 (%)

**Conclusion**

- This is a current study in which a larger number of faecal findings (n=1408) from 6 small rodent species and ferrets were evaluated.
- Compared to the literature<sup>1,3,5</sup>, the frequency of detected parasites was lower in ferrets and either lower or approximately the same in the various small rodents.
- According to the literature<sup>3,4</sup>, the eggs from hamsters, rats and gerbils reported here as *Capillaria*-like eggs could also be *Trichosomoides* eggs (*Trichosomoides crassicauda* in urinary bladder, *Trichosomoides nasalis* in nasal cavity).
- Although the prevalence of *Hymenolepis nana* was much lower than in the literature, rodents represent a potential health risk for humans as *Hymenolepis nana* is a zoonotic parasite. Therefore, faecal examination of small rodents is important to minimize potential risk of human infection.
- The detection of *Giardia* in this study was much lower than in a previous study<sup>3</sup> in which an ELISA was used. Studies have shown that the sensitivity of ELISA is higher compared to microscopic examination for *Giardia*<sup>8</sup>. Therefore, the detection of *Giardia* by ELISA should be preferred in symptomatic rodents and ferrets.

Literature:

- d'Ovidio D, Pepe P, Ianniello D, Noviello E, Quinton JF, Cringoli G, Rinaldi L (2014): First survey of endoparasites in pet ferrets in Italy. Vet Parasitol 203: 227–230.
- d'Ovidio D, Noviello E, Pepe P, Del Prete L, Cringoli G, Rinaldi L (2015): Survey of Hymenolepis spp. in pet rodents in Italy. Parasitol Res 114 (12): 4381–4384.
- Pantchev N, Globokar-Vrhovec M, Beck W (2005): Endoparasitosen bei Kleinsäugetern aus privater Haltung und Igeln. Labordiagnostische Befunde der koprologischen, serologischen und Urinuntersuchung (2002–2004). Tierärztl Prax 33 (K): 296–306.
- Beck w, Pantchev N (2013): Praktische Parasitologie bei Heimtieren. 2.Aufl. Schlütersche, Hannover.
- Kurnosova OP, Arisov MV, Odoyevskaya IM (2019): Intestinal Parasites of Pets and Other House-kept Animals in Moscow. Helminthologia 56 (2): 108–117.
- Bressan MCRV, Calgaro GA, Alexandre SR, Marques T (1997): Prevalence of ecto and endoparasites in mice and rats reared in animal houses. Braz J vet Res anim Sci 34 (3): 142–146.
- Roble GS, Gillespie V, Lipman NS (2012): Infectious disease survey of Mus musculus from pet stores in New York City. J Am Assoc Lab Anim Sci. 51 (1): 37-41.
- Sommer MF, Rupp P, Pietsch M, Kaspar A, Beelitz P (2018) Giardia in a selected population of dogs and cats in Germany - diagnostics, coinfections and assemblages. Vet Parasitol 249:49–56.